In the Specification

Please replace the paragraph beginning on page 41, line 16 with the following amended paragraph:

Liquid cultures are inoculated with a single colony of a polysaccharide-producing strain of bacteria. The preferred strain is designated *Staphylococcus aureus* MN8m, a strain that is a constitutive over-producer of the polysaccharide. A single colony is taken from a tryptic soy agar plate, or similar plate of bacterial growth medium, and grown at 37°C. Temperatures of 10-42°C are also acceptable. Liquid cultures are incubated at 37°C for 1-96 hours while being continuously stirred and flushed with oxygen at a rate of 2 liters/min. The pH is maintained at 7.0 throughout the growth period by the addition of 10 N NaOH via a pH titrator. At the end of the growth period, cell bodies are sedimented at 9000 g for 30 minutes and the supernatant concentrated to ~500 ml via tangential-flow filtration (10,000-500,000 molecular weight cutoff membranes). Two volumes of ethanol are added to precipitate the crude polysaccharide preparation. The precipitate is recovered by centrifugation, re-suspension in water and overnight dialysis against distilled water. The antigen is insoluble. The insoluble, crude antigen is suspended in 50 ml of phosphate buffered saline (PBS, 0.1 M phosphate, 0.15 M sodium chloride) to be digested with the lysozyme (0.5mg) and lysostaphin (0.5 mg) for 0.5 to 16 h at 37°C. Antigen suspensions are further treated with nucleases (0.5 mg) at 37°C for 0.5 to 16 h followed by incubation for 0.5 to 16 h with proteinase K (5 mg) at 37-56°C. After dialysis and lyophilization, dried extracts are dissolved in 5 M HCl and the pH adjusted to 2 with 4 N NaOH. Twenty ml aliquots of this solution are applied to a 5x88 cm column packed with ~~Sephacryl S-300~~ SEPHACRYL S-300® (Pharmacia, Piscataway, NJ) using 0.1 N HCl/0.15 M NaCl buffer with the eluted polysaccharide identified by optical absorption at 206 nm. Fractions corresponding to the polysaccharide representing a continuous range of molecular sizes are separately pooled, dialyzed against water, and lyophilized. Alternately, size fractionation can be performed with a variety of alternative procedures known in the art such as use of diafiltration membranes.

Please replace the paragraph beginning on page 43, line 16 with the following amended paragraph:

Purified dPNAG (10 mg) was dissolved in 0.25 ml of 5 M HCl, neutralized with an equal volume of 5 M NaOH and the final volume adjusted to 2ml with PBS. dPNAG solutions are insoluble at neutral pH but remain completely soluble at slightly acidic or basic pH. Therefore to ensure solubility, the pH of dPNAG solutions was adjusted to 9.0. dPNAG (10mg) was mixed with 1 ml of a 10 mg/ml solution of activated DTm in PBS and pH of the reaction adjusted to 7.5. Two hundred mg of purified sodium cyanoborohydride was added to the mixture and the reaction allowed to proceed in the dark for 14 h at 37°C with mixing. After this time, the reaction mixture was exchanged by dialysis with 0.1 M carbonate buffer, 0.15 M NaCl, pH 10 (10 kDa MWCO dialysis cassette) and the high molecular weight conjugate was purified away from uncoupled components with a ~~Superose 6~~ SUPEROSE 6® prep-grade column by gel filtration chromatography. dPNAG-DTm conjugate was dialyzed against 20 mM HEPES buffer, 50 mM NaCl, pH 8 and stored frozen at –2°C.

Please replace the paragraph beginning on page 44, line 14 with the following amended paragraph:

5 mg of DTm (stock solution in 20 mM HEPES buffer, 50 mM NaCl, pH 8) were dialyzed against borate buffer pH 9.2 for 3h with a 10 kDa MWCO dialysis cassette. After the activation of PNAG with CDAP, 5 mg of DTm was immediately added and the mixture reacted at room temperature for 3h with stirring. After this time, the high molecular weight conjugate was purified from uncoupled components with a ~~Superose 6~~ SUPEROSE 6®prep-grade column by gel filtration chromatography. Fractions containing PNAG-DTm conjugate were pooled, concentrated and stored frozen at –20°C.